SEPARATION MEDIA, MULTIPLE ELECTROSPRAY NOZZLE SYSTEM AND METHOD

[0001] This application claims the benefit of U.S. Provisional Patent Application Ser. No. 60/176,605, filed Jan. 18, 2000, which is herein incorporated by reference in its entirety.

FIELD OF THE INVENTION

[0002] The present invention relates generally to an integrated miniaturized fluidic system fabricated using Micro-ElectroMechanical System (MEMS) technology, particularly to an integrated monolithic microfabricated device capable of generating multiple sprays from a single fluid stream and a separation material, preferably a porous monolithic polymer bed, for chromatographic separations.

BACKGROUND OF THE INVENTION

[0003] New trends in drug discovery and development are creating new demands on analytical techniques. For example, combinatorial chemistry is often employed to discover new lead compounds, or to create variations of a lead compound. Combinatorial chemistry techniques can generate thousands of compounds (combinatorial libraries) in a relatively short time (on the order of days to weeks). Testing such a large number of compounds for biological activity in a timely and efficient manner requires high-throughput screening methods which allow rapid evaluation of the characteristics of each candidate compound.

[0004] The quality of the combinatorial library and the compounds contained therein is used to assess the validity of the biological screening data. Confirmation that the correct molecular weight is identified for each compound or a statistically relevant number of compounds along with a measure of compound purity are two important measures of the quality of a combinatorial library. Compounds can be analytically characterized by removing a portion of solution from each well and injecting the contents into a separation device such as liquid chromatography or capillary electrophoresis instrument coupled to a mass spectrometer.

[0005] Development of viable screening methods for these new targets will often depend on the availability of rapid separation and analysis techniques for analyzing the results of assays. For example, an assay for potential toxic metabolites of a candidate drug would need to identify both the candidate drug and the metabolites of that candidate. An understanding of how a new compound is absorbed in the body and how it is metabolized can enable prediction of the likelihood for an increased therapeutic effect or lack thereof.

[0006] Given the enormous number of new compounds that are being generated daily, an improved system for identifying molecules of potential therapeutic value for drug discovery is also critically needed. Accordingly, there is a critical need for high-throughput screening and identification of compound-target reactions in order to identify potential drug candidates.

[0007] Liquid chromatography (LC) is a well-established analytical method for separating components of a fluid for subsequent analysis and/or identification. Traditionally, liquid chromatography utilizes a separation column, such as a cylindrical tube with dimensions 4.6 mm inner diameter by 25 cm length, filled with tightly packed particles of 5 μm diameter. More recently, particles of 3 μm diameter are being used in shorter length columns. The small particle size provides a large surface area that can be modified with various chemistries creating a stationary phase. A liquid eluent is pumped through the LC column at an optimized flow rate based on the column dimensions and particle size. This liquid eluent is referred to as the mobile phase. A volume of sample is injected into the mobile phase prior to the LC column. The analytes in the sample interact with the stationary phase based on the partition coefficients for each of the analytes. The partition coefficient is defined as the ratio of the time an analyte spends interacting with the stationary phase to the time spent interacting with the mobile phase. The longer an analyte interacts with the stationary phase, the higher the partition coefficient and the longer the analyte is retained on the LC column. The diffusion rate for an analyte through a mobile phase (mobile-phase mass transfer) also affects the partition coefficient. The mobile-phase mass transfer can be rate limiting in the performance of the separation column when it is greater than 2 μm (Knox, J. H. J. *J. Chromatogr. Sci.* 18:453-461 (1980)). Increases in chromatographic separation are achieved when using a smaller particle size as the stationary phase support.

[0008] The purpose of the LC column is to separate analytes such that a unique response for each analyte from a chosen detector can be acquired for a quantitative or qualitative measurement. The ability of a LC column to generate a separation is determined by the dimensions of the column and the particle size supporting the stationary phase. A measure of the ability of LC columns to separate a given analyte is referred to as the theoretical plate number N. The retention time of an analyte can be adjusted by varying the mobile phase composition and the partition coefficient for an analyte. Experimentation and a fundamental understanding of the partition coefficient for a given analyte determine which stationary phase is chosen.

[0009] To increase the throughput of LC analyses requires a reduction in the dimensions of the LC column and the stationary phase particle dimensions. Reducing the length of the LC column from 25 cm to 5 cm will result in a factor of 5 decrease in the retention time for an analyte. At the same time, the theoretical plates are reduced 5-fold. To maintain the theoretical plates of a 25 cm length column packed with 5 μm particles, a 5 cm column would need to be packed with 1 μm particles. However, the use of such small particles results in many technical challenges.

[0010] One of these technical challenges is the backpressure resulting from pushing the mobile phase through each of these columns. The backpressure is a measure of the pressure generated in a separation column due to pumping a mobile phase at a given flow rate through the LC column. For example, the typical backpressure of a 4.6 mm inner diameter by 25 cm length column packed with 5 μm particles generates a backpressure of 100 bar at a flow rate of 1.0 mL/min. A 5 cm column packed with 1 μm particles generates a back pressure 5 times greater than a 25 cm column packed with 5 μm particles. Most commercially available LC pumps are limited to operating pressures less than 400 bar and thus using an LC column with these small particles is not feasible.

[0011] More recently, Frechet in U.S. Pat. Nos. 5,334,310 and 5,453,185, which are each herein incorporated by ref-